

Symmetrical and unsymmetrical analogues of isoxyl; active agents against *Mycobacterium tuberculosis*

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Received 31 May 2006; revised 30 June 2006; accepted 30 June 2006

Available online 27 July 2006

Abstract—Symmetrical and unsymmetrical analogues of the antimycobacterial agent isoxyl have been synthesized and tested against *Mycobacterium tuberculosis* H37Rv and *Mycobacterium bovis* BCG, some showing an increased bactericidal effect. In particular, compounds 1-(*p*-*n*-butylphenyl)-3-(4-propoxy-phenyl) thiourea (**10**) and 1-(*p*-*n*-butylphenyl)-3-(4-*n*-butoxy-phenyl) thiourea (**11**) showed an approximate 10-fold increase in in vitro potency compared to isoxyl, paralleled by increased inhibition of mycolic acid biosynthesis in *M. bovis* BCG. Interestingly, these isoxyl analogues showed relatively poor inhibition of oleate production, suggesting that the modifications have changed the spectrum of biological activity.

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Mycobacterium tuberculosis continues to be one of the most prevalent causes of morbidity and mortality worldwide.¹ The impact of this infection on nearly one-third of the world's population justifies extensive research into new chemotherapeutic reagents.¹ Eisman et al. first published in vitro and in vivo data on the anti-tuberculosis activity of substituted thioureas tested in mice and guinea pigs.² Subsequent modification and biological testing produced a library of compounds, in which the thiourea derivative 4,4'-diisooamylthio-carbanilide (isoxyl, thiocarlide, ISO [**1**]) was shown to have considerable antimycobacterial activity in mice, guinea pigs and rabbits.^{3,4} Clinical use of ISO began in the 1960s to treat tuberculosis.^{5–8} Furthermore, ISO was shown to be effective against various multi-drug resistant *M. tuberculosis* isolates.⁹ In an attempt to understand the biological basis for the activity of ISO, Phetsuksiri et al. investigated its mode of action and reported that ISO, like isoniazid and

ethanionamide, strongly inhibited the synthesis of mycolic acids.¹⁰

The fatty acid synthase I (FAS-I) in mycobacteria synthesizes long-chain fatty acids, including stearic acid, which are subsequently desaturated to oleic acid, a ubiquitous constituent of mycobacterial membrane phospholipids.^{11,12} Over-expression of the *M. tuberculosis* putative fatty acid desaturases in *Mycobacterium bovis* BCG identified *desA3*, which encodes the $\Delta 9$ -acyl-CoA desaturase responsible for the biosynthesis of oleic acid. Over-expression of *desA3* also resulted in increased resistance of *M. bovis* BCG to ISO, identifying *desA3* as a novel target.¹⁰

The crystal structure of the related $\Delta 9$ -stearoyl-acyl carrier protein desaturase has been resolved to 2.4 Å.¹³ By using docking experiments, it has become possible to suggest modifications to the ISO skeleton that may improve the affinity of the inhibitor for the active site. As a result an initial series of symmetrical and unsymmetrical ISO analogues have been developed as potential antimycobacterial agents.

Symmetrical ISO analogues were prepared using a four-step sequence.^{14,15} 4-Acetomidophenol is alkylated upon base treatment with an appropriate halide substituent. The acetamide functionality is deprotected to form the

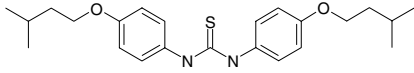
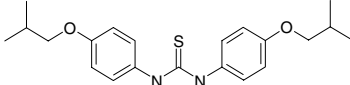
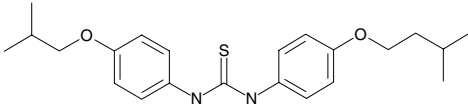
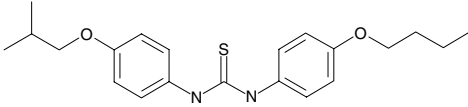
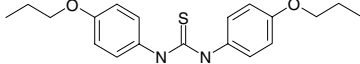
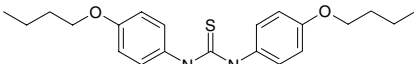
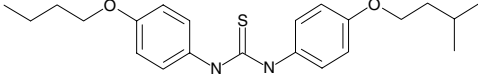
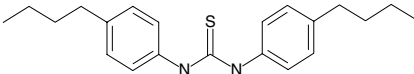
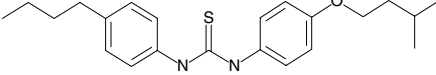
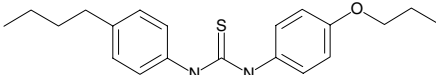
Abbreviations: ACP, acyl carrier protein; CoA, coenzyme A; DTT, dithiothreitol; FAS, fatty acid synthase; FAMES, fatty acid methyl ester; MAMEs, mycolic acid methyl ester; MIC, minimum inhibition concentration; SI, selectivity index.

Keywords: *Mycobacterium tuberculosis*; Mt; *Mycobacterium bovis*; Mb; Mycolic acids; Isoxyl; Inhibitor.

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Table 1. Structures, in vitro and in vivo results of isoxyl analogues^{20–22}

Compound	Structure	Yield	<i>M. tb</i> MIC ₉₉ ^a ($\mu\text{g/ml}$) ¹⁸	<i>M. bovis</i> BCG MIC ₉₉ ^a ($\mu\text{g/ml}$) ¹⁸	FAMES and MAMES			Oleate inhibition ^{e,19}	Cytotoxicity SI ²³
					α^b	Keto ^c	FA ^d		
1		53%	2.0	1.1	60.45	58.63	114.12	61.02	82
2		30%	ND ^f	0.9	46.37	45.83	110.36	51.62	ND ^f
3		55%	0.39	0.9	66.45	42.64	168.36	19.24	72
4		47%	0.1	0.6	56.65	36.50	132.26	63.99	79.9
5		35%	0.2	0.5	49.50	41.95	114.40	90.76	44
6		35%	ND ^f	0.5	43.73	33.92	123.10	94.25	ND ^f
7		55%	<0.1	0.4	61.27	44.05	136.04	79.96	89
8		52%	ND ^f	0.075	62.66	33.98	132.86	NI ^g	ND ^f
9		65%	ND ^f	0.6	38.78	30.51	121.47	14.82	ND ^f
10		85%	<0.1	0.06	53.79	37.40	130.65	NI ^g	100

11		84%	<0.1	0.7	45.51	26.90	118.61	NI ^g	87
12		70%	0.78	0.6	62.25	27.39	104.41	NI ^g	12.4

Table 1 (continued)

Compound	Structure	Yield	<i>M. tb</i> MIC ₉₉ ^a (μg/ml) ¹⁸	<i>M. bovis</i> BCG MIC ₉₉ ^a (μg/ml) ¹⁸	FAMES and MAMES			Oleate inhi- bition ^{e,19}	Cyto- xicity SI ²³
					α ^b	Keto ^c	FA ^d		
13		59%	>6.25	0.7	54.42	35.76	131.38	17.75	ND ^f
14		48%	0.2	0.06	46.50	34.60	142.38	NI ^g	45.5
15		45%	ND ^f	0.5	88.84	66.09	183.79	53.32	ND ^f
16		50%	0.2	1.1	50.79	32.56	147.92	48.83	48
17		57%	0.39	1.5	54.94	39.12	140.70	66.15	22.1
18		48%	1.56	1.75	62.54	41.69	158.83	NI ^g	5.28

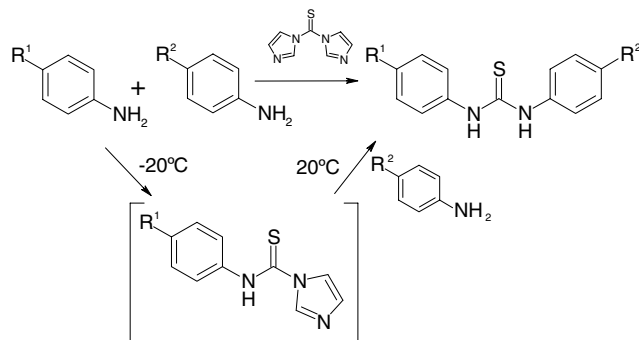
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amine under acidic reflux conditions. The resulting product is coupled using ethanolic carbon disulfide, catalyzed by sulfur to generate the symmetrical ISO analogues. The products were recrystallized and fully characterized by NMR (¹H and ¹³C) and mass spectrometry (1–2 and 5–6) (Table 1).

Unsymmetrical ISO analogues were synthesized by two methods. Isothiocyanate groups can be coupled with amines to generate the isothiurea subunit indicative of ISO analogues (Scheme 1). Commercially available 4-*n*-butylphenyl isothiocyanate was used to generate three novel unsymmetrical ISO analogues



Scheme 1.



Scheme 2.

(9–11) (Table 1). 4-*n*-Butylphenyl isothiocyanate is coupled with an amine substituent in ethanol at 50 °C and the products recrystallized from hot ethanol. These compounds (9–11) were fully characterized by NMR (¹H and ¹³C) and mass spectrometry. More recently, Sriram et al. prepared a series of isonicotinyl hydrazones by reacting isonicotinyl hydrazide (INH) with thiourea units derived from various phenyl isothiocyanate substituents.¹⁶

Due to the lack of commercially available isothiocyanate substituents a second method was developed (Scheme 2). Perkins et al. coupled two different amine substituents with 1,1-thiocarbonyldiimidazole, by controlling the temperature of the reaction.¹⁷ In a one-pot reaction the first equivalent of the amine substituent is reacted with 2 equiv of 1,1-thiocarbonyldiimidazole at –20 °C in acetonitrile. Once the amine has fully reacted, the second equivalent of a different amine is added and the temperature of the reaction is raised to 20 °C. The desired products were recrystallized and fully characterized by NMR (¹H and ¹³C) and mass spectrometry (3, 4, 7 and 13–21) (Table 1).

As shown by Phetsuksiri et al. ISO is an anti-mycobacterial agent implicated in targeting the membrane-bound $\Delta 9$ -desaturase, DesA3 and unknown targets in mycolic acid biosynthesis.¹⁰ Therefore, aryl thioureas present themselves as anti-tuberculosis drugs worthy of further development.

The use of an ISO analogue without functional groups attached to the core thiourea unit (22) clearly showed that, without modification, poor in vivo and in vitro activity was observed; therefore the functional R¹ and R²-groups of ISO play a key role in the effectiveness of the compound. The incorporation of R-group functions (see Table 1) leads to an increase in activity of the compounds and gives justification for functionality at both positions. The use of a biphenyl at one or both of the positions (16–18) was also tolerated in comparison to ISO in terms of in vivo activity. The incorpora-

tion of biphenyl to both positions (19) produced poorer in vivo activity in comparison to ISO (1), however inhibition was shown against, both mycolate and oleate production. Interestingly the unsymmetrical biphenyl ISO (18) failed to inhibit oleate production leading to the possible conclusion that this compound does not inhibit DesA3 desaturase activity but some other enzyme specifically involved in the production of mycolic acids.

Numerous compounds in this study have similar if not better activities than that of ISO (see Table 1). Notably, the use of an aliphatic C₄ functionality to either or both R¹ and R²-positions increased the potency of the inhibitor (4, 7–8, 10–11 and 14). Compound 10 has been shown to be the most effective inhibitor in this study possessing a significant 10-fold increase in potency against both *M. bovis* and *M. tuberculosis* H37Rv. It shows similar effects with regard to inhibition of mycolate production, but unusually does not inhibit oleate production in vitro. This leads to the possibility that compound 10 does not inhibit the $\Delta 9$ -desaturase specifically, but inhibits other enzymes involved in the production of mycolic acids.

Interestingly, the oxygen containing compounds produced in this study indicate that oxygen is required for a more effective inhibitor of oleate biosynthesis. Nearly all the compounds that have one or no oxygens tend to be poor inhibitors of oleate biosynthesis, whereas compounds containing two oxygens (2–7) inhibit oleate biosynthesis, to the same degree if not better than ISO (1). Compounds 5–7 all showed a high degree of activity against the $\Delta 9$ -desaturase inhibitory oleate biosynthesis in vitro 91%, 94% and 80%, respectively.

In conclusion, the analogues in this study have provided an insight into the functionalities and their roles in inhibition of ISO. The new analogues 10 and 11 have all show improved activities against *M. tuberculosis* to such a degree that the MIC values are nearly the same or better than those of other well-known anti-tuberculosis inhibitors, such as isoniazid and rifampin.

Acknowledgments

This work was supported by the Medical Research Council; G.S.B. acknowledges support from Mr. James Bardrick in the form of a Personal Research Chair and as a former Lister Institute Jenner Research Fellow. D.E.M. is a Leverhulme Trust Emeritus Research Fellow. This work was supported by Tuberculosis Antimicrobial Acquisition and Coordinating Facility (TAACF).

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18. *Minimum inhibition concentration (MIC₉₉) and whole cell radiolabelling.* MIC₉₉ of ISO analogues against *M. bovis* BCG were calculated by growth on solid media and whole cell radiolabelling are as described previously.^{9,10}
19. *In vitro effect of ISO on oleic acid synthesis.* The wild type *M. bovis* BCG strain was grown in Sauton medium supplemented with 0.025% tyloxapol. Cells were harvested by centrifugation, resuspended in 0.25 M sucrose, pulse-disrupted by probe sonication and centrifugation at 27,000g for 30 min at 4 °C. The Δ9-stearoyl-CoA desaturase activity was assayed as described.¹⁰
20. *1,3-Bis-(p-2-methylpropoxyphenyl)thiourea.* A solution of 4-(2-methylpropoxy)aniline (500 mg, 3.2 mmol, 1 equiv) in ethanol (50 ml) was treated with carbon disulfide (0.18 ml, 3.2 mmol, 1 equiv) and sulfur (28 mg, 0.8 mmol, 0.25 equiv), and heated under reflux for 16 h. The ethanol was removed in vacuo to yield the crude product as an off-white solid that was recrystallized from methanol, to yield the title compound in 30% yield (100 mg). ¹H NMR (CDCl₃, 300 MHz) δ_H: 0.98 (d, 12H, *J* = 6.6 Hz, CH(CH₃)₂), 2.12 (m, 2H, CH(CH₃)₂), 3.72 (d, 4H, *J* = 6.5 Hz, OCH₂CH), 6.90 (d, 4H, *J* = 8.8 Hz, H-3, H-5), 7.37 (d, 4H, *J* = 8.8 Hz, H-2, H-6), 7.71 (s, 2H, NH); ¹³C NMR (CDCl₃, 75 MHz) δ_C: 21.30 (CH(CH₃)₂), 29.40 (CH(CH₃)₂), 75.9 (OCH₂CH₃), 116.41 (C-3, C-5), 128.5 (C-2, C-6), 135.2 (C-1), 158 (C-4), 181 (NHCSNH) *m/z* (EI) 372 (M⁺ 33%), 109 (MH²⁺-CH₂CH(CH₃)₂ and CSNHPhOCH₂CH(CH₃)₂, 100%); HRMS calcd for C₂₁H₂₈N₂O₂S (MH⁺): 395.1769, found 395.1783; mp 156–158 °C.
21. *1-(p-n-Butylphenyl)-3-(4-n-butoxy-phenyl) thiourea.* 4-n-Butylphenyl isothiourea (1 ml, 5.8 mmol, 1 equiv) was added to ethanol (30 ml) at room temperature before addition of 4-butoxyaniline (0.96 g, 5.8 mmol, 1 equiv). After stirring at 50 °C for 2 h, the reaction mixture is cooled to room temperature and the crude product crashed out. The title compound was recrystallized using hot ethanol to give 84% yield (1.75 g). ¹H NMR (CDCl₃, 300 MHz) δ_H: 0.85 (t, 3H, *J* = 7.4 Hz, OCH₂CH₂CH₂CH₃), 1.05 (t, 3H, *J* = 7.1 Hz, CH₂CH₂CH₂CH₃), 1.25 (m, 2H, CH₂CH₃), 1.30–1.50 (m, 2H, CH₂CH₂CH₃), 1.70–1.85 (m, 2H, OCH₂CH₂CH₂), 2.60 (m, 2H, CH₂CH₂CH₂CH₃), 3.85 (t, 2H, *J* = 6.5 Hz, OCH₂), 6.75 (d, 2H, *J* = 8.4 Hz, H-3', H-5'), 7.15–7.25 (m, 6H, H-2, H-6, H-3, H-5, H-2', H-6'), 7.75 (s, 2H, NH); ¹³C NMR (CDCl₃, 75 MHz) δ_C: 10.2 (CH₂CH₃), 13.6 (CH₂CH₃), 22.0 (CH₂CH₃), 22.2 (CH₂CH₃), 33.2 (CH₂CH₂CH₂), 34.9 (OCH₂CH₂CH₃), 69.5 (OCH₂), 115.0 (C-3', C-5'), 125.1 (C-3, C-5), 127.3 (C-2, C-6), 129.2 (C-2', C-6', C-4), 135.5 (C-1), 141.1 (C-4), 159.2 (C-1'), 180.3 (NHCSNH); *m/z* (EI) 379.4 [M⁺ Na⁺] (75%); HRMS calcd for C₂₁H₂₈N₂O₂S (MH⁺): 354.1653, found 354.1645; mp 137–140 °C.
22. *1-(p-n-Butoxyphenyl)-3-(p-methylbutoxy-phenyl)thiourea.* 1,1-Thiocarbonyldiimidazole (434 mg, 2.42 mmol, 2 equiv) is dissolved in anhydrous acetonitrile and left at –20 °C, while 4-(3-methylbutoxy)aniline (200 mg, 1.2 mmol, 1 equiv) was added dropwise to the solution. The reaction is monitored by TLC until all the aniline substituent is consumed. The second substituent is added, 4-butoxyaniline (200 mg, 1.2 mmol, 1.1 equiv) is then added dropwise and left to come to room temperature gradually. The acetonitrile is reduced in vacuo and the organic layer is extracted with ethyl acetate. This is acidified to pH 2 with 1 M HCl. The organic layer is then washed with water, brine, dried and reduced in vacuo. The compound is recrystallized cold in ethyl acetate in petrol to give the title compound in 55% yield (232 mg). ¹H NMR (CDCl₃, 300 MHz) δ_H: 0.95–1.05 (m, 9 H, CH₃), 1.40 (m, 2H, CH₂CH₃), 1.65 (m, 2H, CH₂CH(CH₃)₂), 1.70–1.80 (m, 3H, CH(CH₃)₂ and CH₂CH₂CH₃), 3.95 (m, 4H, OCH₂), 6.40 (d, 4H, *J* = 6.5 Hz, H-3, H-5), 6.60 (d, 4H, *J* = 6.7 Hz, H-2, H-6), 7.65 (s, 1H, NH); ¹³C NMR (CDCl₃, 75 MHz) δ_C: 14.0 (CH₃), 20.1 (CH₂CH₃), 24.5 (CH(CH₃)₂), 25.7 (CH(CH₃)₂), 35.6 (CH₂CH₂CH₃ and CH₂CH(CH₃)₂), 70.9 (OCH₂), 115.6 (C-3, C-5), 127.8 (C-2, C-6), 131.2 (C-4), 155.6 (C-1), 180.4 (NHCSNH); *m/z* (EI) 409.4 [M⁺ Na⁺] (75%); HRMS calcd for C₂₂H₃₀N₂O₂S (MH⁺): 386.5514, found 386.5569; mp 145–147 °C.
23. Compounds were screened by serial dilution to assess toxicity to a VERO cell line, generally beginning at 10× the MIC. Selectivity index (SI) has defined as the ratio of the measured IC₅₀ in VERO cells to the MIC value.